

Epstein-Barr virus—recent advances

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THE LANCET
Infectious Diseases

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Epstein-Barr virus is a tumorigenic herpes virus that is ubiquitous in the adult population. The virus is generally spread to and between young children through salivary contact, and only causes clinical illness where primary infection is delayed until adolescence or beyond, when an intense immunopathological reaction leads to the symptoms of infectious mononucleosis in roughly 50% of cases. More than 90% of the world's population carry Epstein-Barr virus as a life-long, latent infection of B lymphocytes. Recent data show that by mimicking B-cell antigen-activation pathways the virus enters the long-lived memory B lymphocyte pool where it evades immune elimination by severely restricting its own gene expression. By influencing B-cell survival mechanisms Epstein-Barr virus may induce tumours such as B lymphoproliferative disease and Hodgkin's disease. Vaccines are being developed to prevent and/or treat these conditions, but an animal model is required to study pathogenesis before a rational vaccine strategy can be formulated.

Lancet Infect Dis 2003; **3**: 131–40

Epstein-Barr virus (EBV; figure 1) has co-evolved with human beings over millions of years and during this long association the virus life cycle has become supremely adapted to its human host so that it is one of our most effective parasites. Since its discovery in cultured Burkitt's lymphoma cells in 1964,¹ EBV has been implicated in a wide variety of diseases, both benign and malignant, of either lymphoid or epithelial origin (table 1). Like all herpes viruses, EBV has latent and productive (lytic) phases in its life cycle, the former maintaining the virus long term in its host and the latter effecting virus production and spread. A unique set of latent genes (outlined in table 2) confer EBV with oncogenic potential and the ability to induce immortalisation of B lymphocytes in vitro. Despite this, EBV establishes a harmless life-long infection in almost everyone worldwide and rarely causes disease unless the host-virus balance is upset.

This review concentrates on recent major advances in our understanding of EBV biology and pathogenesis, and addresses topical issues relating to vaccine development for disease prevention. Not all of the EBV-associated tumours are discussed in detail here, but they are listed in tables 1 and 2, and have been reviewed recently.⁶

Epidemiology

More than 90% of adults worldwide have been infected with EBV⁷ and carry the virus as a life-long persistent infection,

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Robert Harding

Figure 1. Colourised transmission electron micrograph of the Epstein-Barr virus, x40 000.

with latent infection of B lymphocytes^{8,9} and virus production into saliva.^{10,11} In most cases primary infection occurs subclinically during childhood,¹² often by spread between family members via salivary contact.^{13,14} Epidemiological studies in the 1970s showed that primary infection occurs early in non-industrialised countries and in low socioeconomic groups,^{7,15} whereas in affluent societies seroconversion may be delayed until adolescence, when infectious mononucleosis (IM) develops in between 50% and 74% of cases.^{16–19} Changing demography and life styles in the intervening 30 years are likely to have altered this pattern, and new studies are required before a logical vaccine strategy can be planned.

It is commonly assumed that those who remain uninfected throughout childhood generally become infected as adolescents through kissing, and consequently IM is often called the kissing disease. However, there are single reports

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Table 1. EBV-associated diseases

Disease	At risk population	EBV association
Lymphocyte origin		
Infectious mononucleosis (IM)	Adolescents/young adults from western societies/ high socioeconomic groups	Majority. IM-like syndromes also occur in cytomegalovirus, HIV primary infection
X-linked lymphoproliferative syndrome (XLPS)	Male offspring of female carriers of XLPS mutation	Majority. A few non-EBV-associated lymphomas occur in children with the mutation
B lymphoproliferative disease (BLPD)	Post-transplant lymphoproliferative disease HIV infection—primary central nervous system lymphoma —peripheral lymphoma	^90% <100% ^50%
Burkitt's lymphoma (BL)	African children—endemic BL HIV infection—sporadic BL	97–100% ^25%
Hodgkin's disease	Children—developing countries Young adults—high socioeconomic groups —history of IM	Overall ^65% Mixed cellularity type 80% Childhood ^80%
T/NK cell lymphoma	Chronic active EBV HIV infection	10–100%, depending on histological type
Primary effusion lymphoma	HIV infection	70–80%. 100% contain HHV8 DNA
Epithelial cell origin		
Oral hairy leukoplakia	HIV infection Other immunodeficiencies	100%
Nasopharyngeal carcinoma	S Chinese and Inuit races—high incidence Mayaks, Dyaks, Indonesians, Filipinos, Vietnamese—moderate incidence	Non-keratinised 100% Keratinised 30–100%
Gastric carcinoma	Not identified	Undifferentiated carcinoma of naso-pharyngeal type 100% Adenocarcinoma 5–15%
Other		
Leiomyosarcoma	HIV infection—mainly children Immunodeficiency—mainly children	Not known
Additional tumours with possible EBV associations		
Salivary gland tumours ²		
Breast carcinoma ³		
Hepatocellular carcinoma ⁴		
Thymoma ⁵		

HHV8=human herpesvirus 8; ^=about.

of EBV detection in male²⁰ and female²¹ genital secretions, suggesting the possibility of sexual transmission. A recent seroepidemiological study on university students lends support to this possibility by showing strong correlations of both EBV seropositivity and history of IM with sexual intercourse and increasing numbers of sexual partners.²² However, these data are not conclusive because they do not differentiate between direct transmission in genital secretions and spread by practices associated with sexual intercourse such as kissing.

Latent EBV infection of B lymphocytes in the blood of healthy donors affords another potential route of transmission; infection has been documented after infusion of a large volume of fresh blood.^{23,24} Transmission from a transplanted organ can also occur with subsequent infection of a previously seronegative recipient,^{25,26} and is a risk factor for post-transplant lymphoproliferative disease (PTLD).

It is now known that there are two types of EBV (1 and 2 or A and B) circulating in the community, which show variation in DNA sequence of the latent genes. They show no specific disease association but type 1 is more prevalent in the west, whereas types 1 and 2 are equally prevalent in Africa and Papua New Guinea.²⁷ Additionally, different EBV isolates can be distinguished by varying lengths of genomic

repeat sequences, and these have been used to track individual isolates through families and in organ donor/recipient pairs.^{14,25,26}

EBV life cycle

Our understanding of the virus infectious cycle is based mainly on observations in acute IM, since studies of symptomless primary infection are not often feasible. EBV infects at mucosal surfaces, but whether epithelial-cell infection is part of the normal virus life cycle remains unresolved. In spite of EBV's known tropism for epithelial cells, exemplified by the several EBV-associated tumours/diseases of epithelial origin (table 1), no EBV-infected epithelial cells have been identified in tonsils removed during IM, whereas both latent and lytic infection of B cells can regularly be detected.²⁸ Since concentrations of EBV in saliva are generally low, it is reasonable to assume that incoming virus from this source requires amplification to establish persistence in the B cell population of its new host. Thus, there could be an initial round of lytic replication in tonsillar epithelium with secondary infection of underlying B cells.^{29,30} However, at present the weight of evidence supports direct infection of B cells in the lymphoepithelium of Waldeyer's ring. Here crypt structures

Table 2. EBV latent antigens

EBV antigen	Required for immortalisation	Function known/postulated	Expressed in vivo in							
			PBM	IM	BL	NPC	HD	BLPD	TCL	Gastric carcinoma
EBNA1	+	Genome maintenance	?	+	+	+	+	+	+	+
EBNA2	+	Viral oncogene, transactivates cellular and other latent viral genes	–	+	–	–	–	+	–	–
EBNA3A	+	Activates cellular genes	–	+	–	–	–	+	–	–
EBNA3B	–	Activates cellular genes	–	+	–	–	–	+	–	–
EBNA3C	+	Viral oncogene, increases LMP1 expression	–	+	–	–	–	+	–	–
EBNA LP	+/-	Co-activates EBNA2 responsive genes, increases efficiency of immortalisation	–	+	–	–	–	+	–	–
LMP1	+	Viral oncogene, induces B cell activation and adhesion, protects from apoptosis	–	+	–	(+)	+	+	(+)	?
LMP2	–	Repression of lytic cycle, enhances B cell survival	+	+	–	+	+	+	+	+

(+)=expressed in a proportion of cases; PBM=peripheral blood mononuclear cells; HD=Hodgkin's lymphoma; EBNA=EBV nuclear antigen; IM=infectious mononucleosis; BLPD=B cell lymphoproliferative disease; LP=leader protein; BL=Burkitt's lymphoma; TCL=T cell lymphoma; LMP=latent membrane protein; NPC=nasopharyngeal carcinoma; ?=not known

that dip into underlying lymphoid tissue could allow EBV direct access to B cells.^{31,32} In this case, expression of the latent viral genes, which induce B cell proliferation, would serve to amplify the number of virus-infected cells in the body and establish latency before immune mechanisms develop.

Detailed analysis of tonsillar B cells suggests that EBV uses physiological B cell antigen-activation pathways to establish and maintain latency.³³ The hypothesis is that newly infected naive tonsillar B cells, under the influence of the oncogene EB viral nuclear antigen (EBNA) 2, express all the latent viral genes, and proliferate in germinal centres. After this stage an alteration in viral gene expression, with downregulation of EBNA 2, halts proliferation and allows differentiation. Normal B cells require two signals for their survival in germinal centres, where most die by apoptosis. For B cells uninfected with EBV, these signals are delivered by binding to helper T cells, via the CD40 molecule present on the B cell surface, and the B cell receptor (BCR), binding to its specific antigen on dendritic cells. Cells infected with EBV are thought to avoid apoptosis in this environment by expressing EBV latent membrane proteins (LMP) 1 and 2a. These molecules together could provide the necessary survival signals because LMP1 is a CD40 homologue and LMP2a mimics BCR engagement.^{34–38} This mechanism would minimise loss of EBV-infected B cells by ensuring their progression into the long-lived memory B cell population, and provide a rationale for the evolutionary conservation of these latent genes. However, although latency in B cells is a successful strategy for survival, it can occasionally allow survival of an abnormal cell with the capacity for uncontrolled proliferation.

After primary infection and the establishment of persistence, EBV can be detected in 0.5–50 in every million B cells in peripheral blood.^{39,40} Here EBV gene expression is restricted, possibly to only LMP2a,^{41,42} a protein that maintains latency by providing the survival signals, and

inhibiting B cell activation and lytic cycle entry.^{43,44} These memory B cells are resting cells with low-level expression of costimulatory molecules, which are thereby assumed to evade immune recognition and elimination.⁴⁵ To complete its life cycle EBV must reactivate from latency and infect other susceptible individuals. The molecular details of the reactivation process are unclear, but it is possible that the trigger for memory B cells to enter the lytic cycle is again physiological. Activation by specific antigen would induce homing of infected memory B cells to the tonsil and maturation to plasma cells, a process that has been linked to lytic cycle entry.⁴⁶ From this site infectious virus may infect co-resident naive B cells or be shed into saliva. Thus, the persistently infected B cell pool is maintained and the virus life cycle completed.

Effective persistence of the virus despite life-long active immune surveillance by the host suggests effective immune evasion. EBV, like other herpes viruses, has evolved sophisticated mechanisms to safeguard its life cycle. As discussed earlier, during latency viral gene expression is severely restricted, although it is not clear which viral proteins (if any) are expressed. However, expression of EBNA1 is essential during cell division when it mediates binding of viral DNA to cellular chromosomes, ensuring its equal partitioning into daughter cells.⁴⁷ This unusual protein contains a glycine/alanine repeat region which confers resistance to degradation by the host cell proteasome.⁴⁸ Thus no endogenous peptides are produced for display on the B cell surface for presentation to CD8 T cells. Although this would seem to be an immune evasion mechanism, it may have evolved simply to stabilise the vital EBNA1 protein at a sufficient intracellular concentration to maintain the EBV genome.⁴⁹ However, there is no doubt that BL cells, in which EBNA1 is the only known viral protein to be expressed, can proliferate in vivo without invoking effective immune control.

When EBV reactivation occurs, several lytic viral proteins are expressed which actively inhibit immune mechanisms. These include an interleukin 10 homologue that inhibits the costimulatory and antigen-presenting functions of monocytes/macrophages,⁵⁰ and several proteins that impair the release of cytokines, particularly interferon (α and γ).^{51,52} In addition, a Bcl2 homologue prolongs cell survival by inhibiting apoptosis.⁵³ These immune evasion mechanisms in effect buy time to allow effective production of infectious virus. However, in healthy individuals a host-virus balance is achieved such that the virus persists and replicates without harming the host.

Infectious mononucleosis

IM, or glandular fever, is one of the commonest causes of prolonged illness in adolescents and young adults in affluent societies. After an incubation period of between 4 and 7 weeks⁵⁴ signs of IM typified by fever, pharyngitis, lymphadenopathy, splenomegaly, and hepatocellular dysfunction, sometimes with frank jaundice, may ensue.⁵⁵ Splenomegaly in IM is often difficult to detect clinically, although one ultrasound scanning study showed enlargement of the spleen in all 29 cases examined.⁵⁶ Rashes, including macular erythema, petechiae, and urticaria occur in about 3% of cases,⁵⁷ and concurrent administration of ampicillin results in rash in about 90% of cases⁵⁸ (calculated from ref 59).

Confirmation of the diagnosis is usually by serological testing. Heterophile antibody is present in 85% of adolescents and adults with IM,^{60,61} but is often absent in young children.⁶² The Monospot is a quick slide test for the detection of heterophile antibodies. Although reasonably specific, positive tests are also seen in other conditions including HIV, lymphoma, systemic lupus erythematosus, rubella, parvovirus, and other viral infections.^{63–66} IgM to the EB viral capsid antigen (VCA) is both more sensitive and specific, and has almost always developed by the time of clinical presentation. Anti-VCA IgM generally persists for about 1–2 months,⁶⁷ and false positives may be caused by rheumatoid factor.⁶⁸ IgG to VCA persists for life. Extended

serology to detect IgG antibody to early antigens (in the acute phase) or EBNA_s (associated with convalescence) may be useful in selected cases.

Death from IM is very rare, although fatal outcomes have resulted from neurological complications, airway obstruction, splenic rupture, myocarditis, cardiac arrhythmia, liver failure, secondary bacterial infection, and thrombocytopenia (table 3).⁵⁵ Prolonged fatigue, hypersomnia, and short-lived depressive disorders are common after IM.⁶⁹ It is not known whether the fatigue could be ameliorated by a programme of rest and exercise implemented early after the resolution of the acute symptoms, although graded exercise and cognitive behavioural therapy are currently thought to be the most effective treatments for chronic fatigue of unknown cause.⁷⁰ High-dose aciclovir reduces virus production in the throat, although it does not significantly alter the duration of individual symptoms.^{71–73} Studies of corticosteroids in IM show amelioration of acute symptoms; however, the risks of prednisolone are only justified in severe disease,^{74,75} for example where there is incipient airway obstruction,⁷⁶ where steroids may reduce the need for surgical intervention to protect the airway.⁷⁷

The lack of efficacy of aciclovir is because symptoms of IM are not directly due to virus infection of B cells, but are immunopathological, resulting from secretion of cytokines by the large numbers of activated cytotoxic (CD8) T lymphocytes (CTL) typically present in peripheral blood and invading the tissues.^{78,79} These T cells, which can account for as many as 44% of peripheral blood CD8+ lymphocytes, are directed against lytic, and to a lesser extent latent, viral epitopes,⁸⁰ and are thought to control the acute infection by eliminating infected B lymphoblasts. In addition, although less well characterised, natural killer cell and CD4+ T cell responses are also generated in IM, and both these cell types can secrete interferon γ and mediate cytotoxicity.

Chronic active EBV infection

Chronic active EBV (CAEBV) infection is a rare condition, distinct from chronic fatigue, that is typified by severe, chronic, or recurrent IM-like symptoms after a well-

documented primary EBV infection in a previously healthy person. The antibody pattern of acute IM is generally retained (high IgG anti-VCA and EA, failure to produce antibodies to EBNA1, and sometimes persistence of anti-VCA IgM). In addition, there is a marked increase of EBV load in peripheral blood, often with infection of T and/or natural killer cells, and evidence of major organ involvement (eg, interstitial pneumonia, bone marrow hypoplasia, uveitis, hepatitis, and splenomegaly).^{81,82} CAEBV has a high morbidity and mortality from hepatic failure, lymphoma, sepsis, or haemophagocytic syndrome. The disease is difficult to treat; bone marrow transplantation⁸³ or adoptive immunotherapy may be helpful.⁸⁴

Table 3. Complications of primary EBV infection

Organ/system	Complication
Liver	Clinical jaundice (5%), abnormal liver function tests (80–90%), fulminant hepatitis (rare)
Respiratory	Respiratory tract obstruction, interstitial pneumonitis (rare)
Neurological*	Encephalitis, acute cerebellar syndrome, aseptic meningitis, Guillain-Barré syndrome, cranial nerve palsy especially VII, transverse myelitis, seizures, mononeuritis, optic neuritis, cerebral haemorrhage
Spleen	Splenic rupture 0.1–0.5% spontaneous or after mild trauma, usually males, splenic infarction
Haematological	Thrombocytopenia, haemolytic anaemia, neutropenia, haemorrhage secondary to mucosal ulceration
Secondary infection	Streptococcal sore throat, sepsis in association with neutropenia
Psychiatric	Depression, anxiety
Renal	Haematuria, interstitial nephritis, glomerulonephritis (rare)
Cardiac	Myocarditis, pericarditis, arrhythmia, electrocardiogram changes
Immunological	Depressed cell mediated immunity

*Pharyngitis, lymphadenopathy may be absent

X-linked lymphoproliferative syndrome

X-linked lymphoproliferative syndrome (XLP or XLP) is a rare, familial, fatal form of IM that has been recognised for almost 30 years.^{85,86} Typically XLP affects young males who are clinically well before primary EBV infection, but when infected most rapidly succumb to fulminant IM. Death results from hepatic necrosis secondary to massive CTL infiltration and cytokine release, aplastic anaemia or pancytopenia, or in some cases superimposed bacterial or fungal infection.^{87,88} Haemophagocytosis is often a prominent feature.^{87,88} Agammaglobulinaemia and/or B cell lymphoma may occur either before EBV infection or later in those that survive IM.⁸⁹ The acute syndrome is very difficult to treat, but etoposide may have some effect.^{88,90,91} Recently a few patients have undergone bone marrow transplantation with a reported success rate of 50%; however the long-term survival rate is unknown.⁹²

The underlying genetic abnormality in XLP was identified in 1998.^{93–95} This discovery rapidly led to development of diagnostic tests and the recognition that the syndrome consists of a spectrum of clinical manifestations encompassing dysgammaglobulinaemias⁹⁶ (including some cases of common variable immunodeficiency),^{97,98} lymphoid vasculitis with aneurysm formation,⁹⁹ and, rarely, non-EBV-related lymphoma.¹⁰⁰

The defective gene product in XLP is a small, src-homology 2 (SH2) domain-containing cytoplasmic protein named SH2D1A or SAP (signalling lymphocytic-activation molecule [SLAM] associated protein). SAP is expressed in T and natural killer cells and is upregulated by cell activation. SAP interacts with several signalling molecules, the best characterised being SLAM (CD150), which is expressed on activated B and T cells and induces a Th1 phenotype with interferon γ secretion.¹⁰¹ SAP binding to SLAM inhibits signal transduction initiated by SLAM, and thereby controls T cell activation. In natural killer cells SAP binds to 2B4 to mediate cytotoxicity, which is impaired in XLP.¹⁰² In XLP it is probable that the functional absence of SAP allows uncontrolled T cell activation and dysregulated cytokine production. However, it is not clear how this relates to EBV because activation of the SAP/SLAM pathway is not specific to primary EBV infection (H Williams, University of Edinburgh, personal communication).

XLP patients are a difficult group to study, and results of the limited number of immunological investigations undertaken before EBV infection are contradictory, some showing mild immune abnormalities but others not. Combined B and T cell defects are invariably present after EBV infection, although no consistent abnormalities in immune control of EBV have been reported.¹⁰³ Since it is now clear that lymphoma can result from a mutation in the SAP gene in the absence of EBV infection, it is hypothesised that XLP is a more generalised immunodeficiency that can be initiated by several virus infections, rather than an abnormality restricted to immune regulation of EBV. However, the exact nature of the immune deficit remains unknown.

B cell lymphoproliferative disease

EBV-associated B cell lymphoproliferative disease (BLPD) occurs in immunodeficiency states with impaired T cell immunosurveillance, where a lack of EBV-specific CTLs allows proliferation of latently infected B cells.^{104,105} This is a common life-threatening complication of transplantation when immunosuppressive drugs are used to prevent graft rejection. In the context of organ transplantation, BLPD is often known as PTLD. Risk factors for PTLD include high levels of immunosuppressive drugs and primary EBV infection during therapy. Clinical presentations are varied and can mimic graft-versus-host disease, graft rejection, or more conventional infections. Presenting features may resemble an IM-like illness or an extra-nodal tumour, commonly involving the gut, brain, or the transplanted organ.¹⁰⁶ IM-like presentations typically occur in children, within the first year of transplant, and are often associated with primary EBV infection after transfer of donor virus from the grafted organ.^{26,107,108} By contrast, extra-nodal tumours are commonly seen in EBV seropositive recipients several years after the transplant. These varied presentations make the diagnosis of PTLD difficult and consequently predictive markers have been sought. Many studies have investigated EBV load in this context,¹⁰⁹ and high concentrations of EBV DNA in peripheral blood have often been reported in PTLD. However, large fluctuations in viral load are common, and concentrations as high as those seen in PTLD can be seen in healthy transplant recipients.^{110–113} At present there is no standardisation of the technique for viral load estimation between different laboratories; however, with the availability of new rapid, quantitative techniques, continuous monitoring is feasible and may give a clearer indication of those at risk.

PTLD arises as an opportunistic tumour in the setting of intense T cell immune suppression, which allows latently infected B cells to survive and proliferate *in vivo*. Tumour cells generally express all the latent viral genes, and this pattern is assumed to be sufficient to induce a tumorigenic phenotype. Early lesions are often polyclonal with later progression to a monoclonal lymphoma.¹¹⁴ However, the fact that lesions are commonly single suggests that only rare cells have the capacity to form tumours, and the predilection for certain body sites possibly indicates a requirement for specific external stimuli, such as cytokines. No consistent genetic abnormalities have been reported, but recently examination of immunoglobulin heavy chain gene rearrangements in these tumours shows that they often arise from post-germinal-centre B cells with non-functional immunoglobulin gene rearrangements.¹¹⁵ Such cells would normally be eliminated in germinal centres, but are probably rescued by EBV infection, as a consequence of the ability of EBV to affect B cell survival and maturation processes.

First-line treatment for PTLD is reduction of immunosuppressive therapy, which allows recovery of CTL activity and leads to tumour regression in most cases, but risks rejection of the transplanted organ.¹¹⁶ However, PTLD often recurs, and becomes progressively resistant to this conservative form of treatment. Despite the use of conventional lymphoma chemotherapies, the death rate remains at over 50%.¹¹⁷ Novel forms of immunotherapy have

been tested in PTLD, including both antibody and cell-mediated approaches.

Rituximab is a humanised mouse monoclonal antibody that targets the CD20 molecule on the surface of all mature B cells.¹¹⁸ It is currently licensed only for treatment of CD20-positive diffuse large-cell non-Hodgkin lymphoma (NHL), in conjunction with chemotherapy or alone for follicular lymphoma if chemotherapy has failed. Use in PTLD has shown some favourable outcomes^{119,120} although tumour recurrences have been reported.¹²¹ A cytokine release/tumour lysis syndrome has been associated with some fatalities.¹²² Additionally, some instances of fulminant infections have been reported, although whether there is any causal association is unknown.^{121,123,124} At present results are anecdotal and controlled trials are needed to assess the long-term outcome.

Infusions of cultured EBV-specific CTL have been used effectively for both prevention and treatment of PTLD in bone marrow transplant recipients where the donor is available to provide HLA-matched T cells.¹²⁵ However, the production of these donor CTL for individual patients is too expensive and time-consuming for wide application. A related strategy is to develop a bank of HLA-typed CTL that can be used to treat PTLD patients on a best-HLA-match basis. Encouraging results from a pilot study suggest that this may be a more feasible option.¹²⁶ A controlled trial is now underway, and if successful, this form of therapy could be applicable for other infections and tumours in the immunocompromised setting.

Hodgkin's disease

A causative association between EBV and Hodgkin's disease has long been suspected on the grounds of raised concentrations of antibodies to EBV antigens for months or years before onset of Hodgkin's disease,^{127,128} and the increased incidence of Hodgkin's disease in the 5 years following IM.¹²⁹

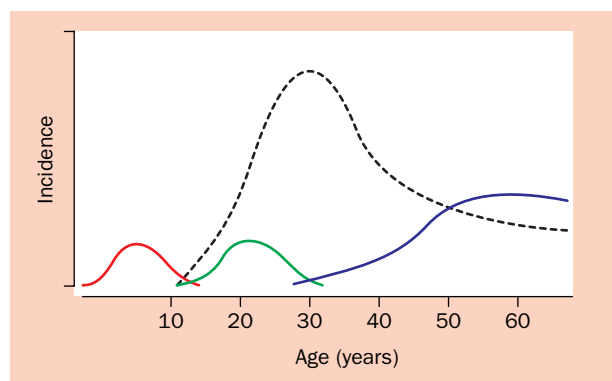


Figure 2. Incidence of Hodgkin's disease by age. Red line=EBV-associated cases; green line=non-EBV-associated cases. A four-disease model is postulated, three peaks are associated with EBV infection—(1) Childhood cases associated with primary infection. Peak age <10 years; this accounts for most childhood cases in developing countries. (2) Young adult age group, 15–34 years associated with delayed primary EBV-infection. (3) Older adult group, accounts for most cases over 55 years. (4) Non-EBV-associated Hodgkin's disease, this is the largest group and accounts for the young adult peak in developed countries. The height of each peak will vary depending on the country in which the data are generated. Based on data included in reference 131. Courtesy of Ruth F Jarrett.

A firm association was finally established by the identification of EBV DNA in the malignant Hodgkin and Reed Sternberg (HRS) cells, albeit in a subset of Hodgkin's disease tumours.¹³⁰ Overall 65% of Hodgkin's disease tumours contain EBV DNA, with the mixed cellularity type being most commonly EBV-associated (table 1).

The age of onset of Hodgkin's disease typically shows a bimodal distribution with one peak in childhood and another at over 50 years of age; however, this varies according to geographical location. The childhood peak occurs later in affluent (15–35 years) than non-affluent (5–10 years) societies. This age difference is reminiscent of primary EBV infection and development of IM, which suggests that Hodgkin's disease might be a non-typical response to primary EBV infection. Although the findings in non-affluent societies seemed to fit this hypothesis, with most Hodgkin's disease developing before the age of 10 being EBV-associated, this is not the case in more affluent societies. Surprisingly, here non-EBV-associated, nodular sclerosing Hodgkin's disease predominates in the adolescent/young adult peak. A recent population-based study in the UK showed non-EBV-associated cases having a unimodal age distribution as above, and a flatter, but probably bimodal, age distribution for the EBV-positive cases (figure 2).¹³¹ It is generally accepted that EBV has an causative role in the pathogenesis of Hodgkin's disease, although the exact mechanistic details are still unclear.

Characterisation of the viral gene expression in HRS cells showed a restricted form of latency with EBNA1, LMP1, and LMP2A expression (figure 3).¹³² LMP1 is known to induce cellular activation and proliferation pathways and inhibit apoptosis, and LMP2 expression enhances cell survival and inhibits lytic cycle activation.¹³³ Viral monoclonality in HRS cells¹³⁴ suggests that EBV infection is an early event in tumour development, and taken together these findings are consistent with an oncogenic role for EBV.

The cellular origin of HRS cells has long been controversial, but recently this has been definitively identified as germinal centre B cells that contain functional immunoglobulin gene rearrangements but show defective immunoglobulin transcription.¹³⁵ This finding, like that in PTLD, suggests that survival of these atypical cells may be an accident of the EBV life cycle in B lymphocytes. The fact that HRS-like cells have also been found in IM lymph nodes¹³⁶ and PTLD¹³⁷ draws parallels between IM and these tumours.

The finding of a viral cause for a subset of Hodgkin's disease tumours opens up the possibility for immunotherapy directed at viral antigens displayed on the tumour cell surface. Although LMP1 and 2 do not contain immunodominant T cell epitopes,¹³⁸ pilot studies are underway to assess CTL therapy for Hodgkin's disease in cases that do not respond to conventional chemotherapy.¹³⁹

EBV in HIV-infected people

Most HIV-infected people are persistently infected with EBV, and with progressive immunodeficiency numbers of EBV-infected B cells in the circulation increase¹⁴⁰ and opportunistic lymphomas may develop. The risk of NHL is increased 60-fold compared with the general population,¹⁴¹ and its presence is an AIDS-defining illness. However, these NHL are not as

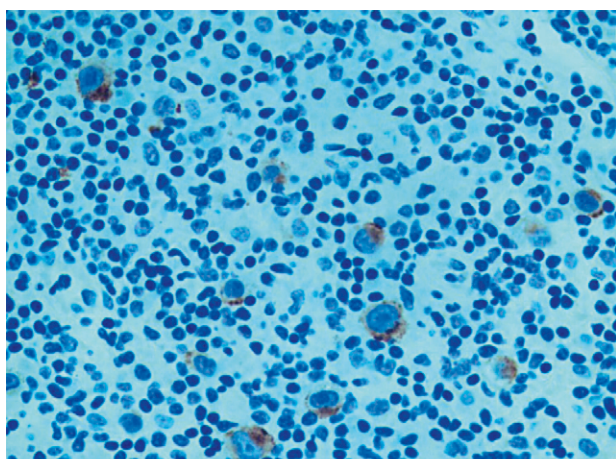


Figure 3. Photomicrograph of mixed cellularity Hodgkin's disease stained for LMP1 expression. There is strongly positive staining in the malignant Hodgkin cells (original magnification $\times 400$). Courtesy of C Bellamy.

uniform as PTLD in their EBV-association and histological type.

Three principal types of NHL are recognised in the HIV setting (table 1), with Burkitt-like tumours developing relatively early in the disease process, whereas primary central nervous system lymphoma (PCNSL) and peripheral NHL occur at a late stage. All three types may be EBV-associated, the strongest association being with PCNSL where almost 100% of tumours contain EBV DNA and express latent viral genes.¹⁴² EBV DNA can be detected in the cerebrospinal fluid in most of these cases, occasionally preceding a visible mass on imaging and forming the basis of a useful diagnostic test.¹⁴³ The incidence of NHL in HIV, particularly that of PCNSL, has fallen since the introduction of highly active antiretroviral therapy (HAART)¹⁴⁴ and there is some evidence that introducing effective antiretroviral therapy prolongs survival after tumour diagnosis.¹⁴⁵

Hodgkin's disease, although not an AIDS-defining illness, is also more prevalent in people with HIV infection, and in this setting EBV-associated mixed cellularity and lymphocyte depleted types are most common.^{146,147}

In addition to lymphomas, EBV is also associated with oral hairy leukoplakia (OHL), which is common in late HIV infection. This is a painless, white, corrugated lesion that was first recognised on the lateral margins of the tongue in AIDS patients,¹⁴⁸ but has now been seen in other immunosuppressed groups.^{149–151} The lesion contains EBV replicating in squamous epithelial cells but without malignant transformation or the establishment of latency.^{152,153} It is unclear whether this represents an amplification of the normal EBV life cycle or infection of an aberrant cell type. OHL is usually symptomless, but can be treated with aciclovir if required.¹⁵⁴

Prospects for a vaccine

At the recent International Workshop on EBV and associated Diseases (Cairns, Australia, July 2002), a discussion session was dedicated to EBV vaccines. After many years in production, there are now two candidate vaccines ready for trial, but there was no consensus on how and when these

should be used. As often happens when commercial companies are involved, there is a conflict between financial and health priorities. Whereas scientists and clinicians believe that EBV-associated tumours in developing nations (nasopharyngeal carcinoma in China, Burkitt's lymphoma in Africa) should be the main priorities, a financially viable vaccine would have to be used in the west for prevention of non-life-threatening cases of IM. Other minor markets would include XLP families and seronegative transplant recipients.

The predominant EBV envelope glycoprotein gp340 has been developed as a vaccine because of its ability to induce neutralising antibody.¹⁵⁵ However, it is unlikely that sterile immunity, which may be necessary for prevention of EBV-induced tumours, will be elicited with this vaccine. Nevertheless, it is possible that this vaccine could prevent IM by moderating the initial viral replication and spread during primary infection, thereby curtailing the massive CTL response to lytic antigens that invokes the immunopathology giving rise to symptoms.

Using our now extensive knowledge of EBV-specific CTL epitopes, a peptide-based vaccine has recently been designed to induce specific cellular responses.¹⁵⁶ To overcome the problem of HLA specificity of viral CTL responses, the extensive allelic variation of human HLA genes, and variation in epitopes between EBV strains, a "polytope" vaccine containing multiple epitopes to which over 94% of the population should respond has been formulated.¹⁵⁷ Again, this vaccine is not designed to induce sterile immunity but it is hoped that it will be effective in the prevention of disease states. An alternative exciting future approach would be to generate therapeutic vaccines in the form of genetically engineered constructs designed to generate or enhance specific immune responses to the latent viral gene products known to be expressed in individual EBV-associated tumours.

Of the many unresolved issues relating to a vaccine for IM, our lack of information on susceptibility remains critical. EBV-seronegative teenagers form the susceptible population, but only around 50% of susceptible people would naturally develop symptoms of IM on seroconversion. Furthermore, without up-to-date studies in a range of social groups and geographical locations, it is not clear how large this population is.

The rationale for prevention of symptoms in IM by neutralising incoming virus and/or reducing the replication of virus after infection depends on the assumption that initial viral dose determines the severity of the disease. This was not

Search strategy and selection criteria

The data used in this review were researched using references from recent articles, as well as abstracts from and presentations at relevant research meetings (particularly the Tumour Associated Herpesviruses 10th Symposium on EBV and Associated Diseases). Medline searches for the year 2001–2002 were undertaken using the following search terms: "Epstein-Barr virus", "infectious mononucleosis", "Hodgkin's disease/lymphoma", "post transplant lymphoproliferative disease", "human immunodeficiency virus (HIV) and Epstein-Barr virus", "oral hairy leukoplakia", "rituximab", and "T lymphocytes and Epstein-Barr virus".

upheld by a recent study, which showed viral loads of equal magnitude in IM and two out of three cases of subclinical seroconversion.¹⁵⁸

The vaccination of healthy individuals against an infection not perceived as being serious demands a very safe and effective vaccine. Therefore, updated epidemiological data and more basic research on pathogenesis of IM are needed, and for this a good animal model is urgently required. The cotton-topped tamarin has been useful in the past for studies on tumour prevention, but is not a good model for IM. The recently discovered EBV-like lymphocryptovirus, naturally endemic in Rhesus monkeys, looks more promising, since

oral infection induces an IM-like syndrome, followed by latent infection in B cells and oropharyngeal secretion.¹⁵⁹ The tumorigenic potential of the virus is under investigation, but it is hoped that experiments in this animal model will assist in unravelling the intricacies of EBV infection in health and disease.

Acknowledgments

We thank Ruth Jarrett at the University of Glasgow for providing figure two, which illustrates a model based on her original data. We also thank Chris Bellamy at the University of Edinburgh for providing figure three.

Conflicts of interest

There are no conflicts of financial, personal, or other interest.

References

- Epstein MA, Achong BG, Barr YM. Virus particles in cultured lymphoblasts from Burkitt's lymphoma. *Lancet* 1964; **1**: 702–03.
- Raab-Traub N, Rajadurai P, Flynn K, Lanier AP. Epstein-Barr virus infection in carcinoma of the salivary gland. *J Virol* 1991; **65**: 7032–36.
- Bonnet M, Guinebretiere J-M, Kremmer E, et al. Detection of Epstein-Barr virus in invasive breast cancers. *J Natl Cancer Inst* 1999; **91**: 1376–81.
- Sugarawa Y, Mizugaki Y, Uchida T, et al. Detection of Epstein-Barr virus (EBV) in hepatocellular carcinoma tissue: a novel EBV latency characterized by the absence of EBV-encoded small RNA expression. *Virology* 1999; **256**: 196–202.
- Leywraz S, Henle W, Chahinian AP, et al. Association of Epstein-Barr virus with thymic carcinoma. *New Engl J Med* 1985; **213**: 1296–99.
- Crawford DH. Biology and disease associations of Epstein-Barr virus. *Phil Trans R Soc Lond B* 2001; **356**: 461–73.
- Henle G, Henle W, Clifford P, et al. Antibodies to Epstein-Barr virus in Burkitt's lymphoma and control groups. *J Nat Cancer Inst* 1969; **43**: 1147–57.
- Babcock GJ, Decker LL, Volk M, Thorley-Lawson DA. EBV persistence in memory B cells in vivo. *Immunity* 1998; **9**: 395–404.
- Thorley-Lawson DA, Miyashita EM, Khan G. Epstein-Barr virus and the B cell: that's all it takes. *Trends Microbiol* 1996; **4**: 204–08.
- Yao QY, Rickinson AB, Epstein MA. A re-examination of the Epstein-Barr virus carrier state in healthy seropositive individuals. *Int J Cancer* 1985; **35**: 35–42.
- Niederman JC, Miller G, Pearson HA, Pagano JS, Dowlatbadi JM. Infectious mononucleosis. Epstein-Barr-virus shedding in saliva and the oropharynx. *N Engl J Med* 1976; **29**: 1355–59.
- Fleisher G, Henle W, Henle G, Lennette ET, Biggar RJ. Primary infection with Epstein-Barr virus in infants in the United States: clinical and serologic observations. *J Infect Dis* 1979; **139**: 553–58.
- Fleisher GR, Pasquariello. Intrafamilial transmission of Epstein-Barr virus infections. *J Pediatr* 1981; **98**: 16–19.
- Gratama JW, Oosterveer MAP, Klein G, Ernberg I. EBNA size polymorphism can be used to trace Epstein-Barr virus spread within families. *J Virol* 1990; **64**: 4703–08.
- Biggar RJ, Henle G, Böcker J, et al. Primary Epstein-Barr virus infections in African infants. II. Clinical and serological observations during seroconversion. *Int J Cancer* 1978; **22**: 244–50.
- Hallee TJ, Evans AS, Niederman JC, Brooks CM, Vogtly JH. Infectious mononucleosis at the United States Military Academy. A prospective study of a single class over four years. *Yale J Biol Med* 1974; **3**: 182–95.
- Infectious mononucleosis and its relationship to EB virus antibody. A joint investigation by university health physicians and PHLS laboratories. *BMJ* 1971; **4**: 643–46.
- Niederman JC, Evans AS, Subrahmanyam L, McCollum RW. Prevalence, incidence and persistence of EB virus antibody in young adults. *N Engl J Med* 1970; **282**: 361–65.
- Sawyer RN, Evans AS, Niederman JC, McCollum RW. Prospective studies of a group of Yale University freshmen. I. Occurrence of infectious mononucleosis. *J Infect Dis* 1971; **123**: 263–70.
- Israele V, Shirley P, Sixbey JW. Excretion of the Epstein-Barr virus from the genital tract of men. *J Infect Dis* 1991; **163**: 1341–43.
- Sixbey JW, Lemon SM, Pagano JS. A second site for Epstein-Barr virus shedding: the uterine cervix. *Lancet* 1986; **2**: 1122–24.
- Crawford DH, Swerdlow AJ, Higgins C, et al. Sexual history and Epstein-Barr virus infection. *J Infect Dis* 2002; **186**: 731–36.
- Gerber P, Walsh JH, Rosenblum EN, Purcell RH. Association of EB-virus infection with the post-perfusion syndrome. *Lancet* 1969; **1**: 593–96.
- Alfieri C, Tanner J, Carpentier L, et al. Epstein-Barr virus transmission from a blood donor to an organ transplant recipient with recovery of the same virus strain from the recipient's blood and oropharynx. *Blood* 1996; **87**: 1122–17.
- Cen H, Breinig MC, Atchison RW, Ho M, McKnight JLC. Epstein-Barr virus transmission via the donor organs in solid organ transplantation: polymerase chain reaction and restriction fragment length polymorphism analysis of IR2, IR3 and IR4. *J Virol* 1991; **65**: 976–80.
- Haque T, Thomas JA, Falk KI, et al. Transmission of donor Epstein-Barr virus (EBV) in transplant organs causes lymphoproliferative disease in EBV-seronegative recipients. *J Gen Virol* 1996; **77**: 1169–72.
- Zimber U, Adldinger HK, Lenoir GM, et al. Geographical prevalence of two types of Epstein-Barr virus. *Virology* 1986; **154**: 56–66.
- Anagnostopoulos I, Hummel M, Kreschel C, Stein H. Morphology, immunophenotype, and distribution of latently and/or productively Epstein-Barr virus-infected cells in acute infectious mononucleosis: implications for the interindividual infection route of Epstein-Barr virus. *Blood* 1995; **85**: 744–50.
- Farrell PJ. Cell-switching and kissing. *Nat Med* 2002; **8**: 559–60.
- Borza CM, Hutt-Fletcher LM. Alternate replication in B cells and epithelial cells switches tropism of Epstein-Barr virus. *Nat Med* 2002; **8**: 594–99.
- Faulkner GC, Krajewski AS, Crawford DH. The ins and outs of EBV infection. *Trends Microbiol* 2000; **8**: 185–89.
- Perry ME, Jones MM, Mustafa Y. Structure of the crypt epithelium in human palatine tonsils. *Acta Otolaryngol* 1988; **454** (suppl): 53–59.
- Babcock GJ, Thorley-Lawson DA. Tonsillar memory B cells, latently infected with Epstein-Barr virus, express the restricted pattern of latent genes previously found only in Epstein-Barr virus-associated tumors. *Proc Natl Acad Sci USA* 2000; **97**: 12250–55.
- Gires O, Zimber-Strobl U, Gonnella R, et al. Latent membrane protein 1 of Epstein-Barr virus mimics a constitutively active receptor molecule. *EMBO J* 1997; **16**: 6131–40.
- Kilger E, Kieser A, Baumann M, Hammerschmidt W. Epstein-Barr virus-mediated B-cell proliferation is dependent upon latent membrane protein 1, which stimulates an activated CD40 receptor. *EMBO J* 1998; **17**: 1700–09.
- Zimber-Strobl U, Kempkes B, Marschall G, et al. Epstein-Barr virus latent membrane protein 1 (LMP1) is not sufficient to maintain proliferation of B cells but both it and activated CD40 can prolong their survival. *EMBO J* 1996; **15**: 7070–78.
- Uchida J, Yasui T, Takaoka-Shichijo Y, et al. Mimicry of CD40 signals by Epstein-Barr virus LMP1 in B lymphocyte responses. *Science* 1999; **286**: 300–03.
- Caldwell RG, Wilson JB, Anderson SJ, Longnecker R. Epstein-Barr virus LMP2A drives B cell development and survival in the absence of normal B cell receptor signals. *Immunity* 1998; **9**: 405–11.
- Miyashita EM, Yang B, Lam KMC, Crawford DH, Thorley-Lawson DA. A novel form of Epstein-Barr virus latency in normal B cells in vivo. *Cell* 1995; **80**: 593–601.
- Wagner HJ, Bein G, Bitsch A, Kirchner H. Detection and quantification of latently infected B lymphocytes in Epstein-Barr virus-seropositive, healthy individuals by polymerase chain reaction. *J Clin Microbiol* 1992; **30**: 2826–29.
- Qu L, Rowe DT. Epstein-Barr virus latent gene expression in uncultured peripheral blood lymphocytes. *J Virol* 1992; **66**: 3715–24.
- Miyashita EM, Yang B, Babcock GJ, Thorley-Lawson DA. Identification of the site of Epstein-Barr virus persistence in vivo as a resting cell. *J Virol* 1997; **71**: 4882–91.
- Miller CL, Burkhardt AL, Lee JH, et al. Integral membrane protein 2 of Epstein-Barr virus regulates reactivation from latency through dominant negative effects on protein-tyrosine kinases. *Immunity* 1995; **2**: 155–66.
- Miller CL, Lee JH, Kieff E, Longnecker R. An integral membrane protein (LMP2) blocks reactivation of Epstein-Barr virus from latency following surface immunoglobulin crosslinking. *Proc Natl Acad Sci USA* 1994; **91**: 772–76.
- Thorley-Lawson DA. Epstein-Barr virus: exploiting the immune system. *Nat Rev Immunol* 2001; **1**: 75–82.
- Crawford DH, Ando I. EB virus induction is associated with B-cell maturation. *Immunology* 1986; **59**: 405–09.
- Yates JL, Warren N, Sugden B. Stable replication of plasmids derived from Epstein-Barr virus in various mammalian cells. *Nature* 1985; **313**: 812–15.
- Levitskaya J, Shapiro A, Leonchik A, Ciechanover A, Masucci MG. Inhibition of ubiquitin/proteasome-dependent protein degradation by the Gly-Ala repeat domain of the Epstein-Barr virus nuclear antigen 1. *Proc Natl Acad Sci USA* 1997; **94**: 12616–21.
- Rickinson AB, Kieff E. Epstein-Barr virus. In: Knipe DM, et al. *Fields virology* 4th edition. Philadelphia: Lippincott Williams & Wilkins, 2001: 2575–627.
- Moore KW, Waal Malefyt R de, Coffman RL, O'Garra A. Interleukin-10 and the interleukin-10 receptor. *Ann Rev Immunol* 2001; **19**: 683–765.
- Cohen JL, Lekstrom K. Epstein-Barr virus BARF1 protein is dispensable for B-cell transformation and inhibits alpha interferon secretion from mononuclear cells. *J Virol* 1999; **73**: 7627–32.
- Morrison TE, Mauser A, Wong A, Ting JP-Y, Kenney SC. Inhibition of IFN- γ signaling by an Epstein-Barr virus immediate-early protein. *Immunity* 2001; **15**: 777–99.
- Henderson S, Huen D, Rowe M, et al. Epstein-Barr virus-coded BHRF1 protein, a viral homologue of Bcl-2, protects human B cells from programmed cell death. *Proc Natl Acad Sci USA* 1993; **90**: 8479–83.
- Hoagland RJ. The incubation period of infectious mononucleosis. *Am J Public Health* 1964; **54**: 1699–705.
- Schooley RT. Epstein-Barr virus (infectious mononucleosis). In: Mandell GL, Bennett JE, Dolin R, eds. *Principles and practice of infectious diseases* 5th edition. Philadelphia, USA: Churchill Livingstone, 2000: 1599–613.
- Dommerby H, Stangerup SE, Stangerup M, Hancke S. Hepatosplenomegaly in infectious mononucleosis, assessed by ultrasonic scanning. *J Laryngol Otol* 1986; **100**: 573–79.

- 57 McCarthy JT, Hoagland RJ. Cutaneous manifestations of infectious mononucleosis. *JAMA* 1964; **187**: 153–54.
- 58 Pullen H, Wright N, Murdoch JM. Hypersensitivity reactions to antibacterial drugs in infectious mononucleosis. *Lancet* 1967; **2**: 1176–78.
- 59 Kerns DL, Shira JE, Go S, et al. Ampicillin rash in children. Relationship to penicillin allergy and infectious mononucleosis. *Am J Dis Child* 1973; **125**: 187–90.
- 60 Evans AS. Infectious mononucleosis and related syndromes. *Am J Med Sci* 1978; **276**: 325–39.
- 61 Fleisher GR, Collins M, Fager S. Limitations of available tests for diagnosis of infectious mononucleosis. *J Clin Microbiol* 1983; **17**: 619–24.
- 62 Sumaya CV, Ench Y. Epstein-Barr virus infectious mononucleosis in children. II. Heterophil antibody and viral-specific responses. *Pediatrics* 1985; **75**: 1011–19.
- 63 Vidrih JA, Walensky RP, Sax PE, Freedberg KA. Positive Epstein-Barr virus heterophile antibody tests in patients with primary human immunodeficiency virus infection. *Am J Med* 2001; **111**: 192–94.
- 64 Horowitz A, Henle W, Henle G, et al. Persistent falsely positive rapid tests for infectious mononucleosis. *Am J Clin Pathol* 1979; **72**: 807–11.
- 65 Wynne Jones J, Pether JVS, Frost RWP. Human parvovirus B19. Hard to differentiate from infectious mononucleosis. London: BMJ Publishing, 1994: 308.
- 66 Hendry BM, Longmore JM. Systemic lupus erythematosus presenting as infectious mononucleosis with a false positive monospot test. *Lancet* 1982; **1**: 455.
- 67 Okano M, Thiele GM, Davis JR, Grierson HL, Purtillo DT. Epstein-Barr virus and human diseases: recent advances in diagnosis. *Clin Microbiol Rev* 1988; **1**: 300–12.
- 68 Henle G, Lennette ET, Alspaugh MA, Henle W. Rheumatoid factor as a cause of positive reactions in tests for Epstein-Barr virus specific IgM antibodies. *Clin Exper Immunol* 1979; **36**: 415–22.
- 69 White PD, Thomas JM, Amess J, et al. Incidence, risk and prognosis of acute and chronic fatigue syndromes and psychiatric disorders after glandular fever. *Br J Psychiatry* 1998; **173**: 475–81.
- 70 Whitting P, Bagnall A-M, Sowden AJ, et al. Interventions for the treatment and management of chronic fatigue syndrome. A systematic review. *JAMA* 2001; **286**: 1360–68.
- 71 Pagano JS, Sixbey JW, Lin J-C. Acyclovir and Epstein-Barr virus infection. *J Antimicrob Chemother* 1983; **12** (suppl): 113–21.
- 72 Andersson J, Britton S, Ernerb J, et al. Effect of acyclovir on infectious mononucleosis: a double-blind, placebo-controlled study. *J Infect Dis* 1986; **153**: 283–90.
- 73 Andersson J, Sködenberg B, Henle W, et al. Acyclovir treatment in infectious mononucleosis: a clinical and virological study. *Infection* 1987; **15**: S14–20.
- 74 Brandfonbrener A, Epstein A, Wu S, Phair J. Corticosteroid therapy in Epstein-Barr virus infection. Effect on lymphocyte class, subset, and response to early antigen. *Arch Intern Med* 1986; **146**: 337–39.
- 75 Straus SE, Cohen JI, Tosato G, Meier J. Epstein-Barr virus infections: biology, pathogenesis, and management. *Ann Intern Med* 1993; **118**: 45–58.
- 76 McGowan JE Jr, Chesney PJ, Crossley KB, LaForce FM. Guidelines for the use of systemic glucocorticosteroids in the management of selected infections. *J Infect Dis* 1992; **165**: 1–13.
- 77 Suderick RM, Narula AA. Steroids for airway problems in glandular fever. *J Laryngol Otol* 1987; **101**: 673–75.
- 78 Foss H-D, Herbst H, Hummel M, et al. Patterns of cytokine gene expression in infectious mononucleosis. *Blood* 1994; **83**: 707–12.
- 79 Linde A, Andersson B, Svenson SB, et al. Serum levels of lymphokines and soluble cellular receptors in primary Epstein-Barr virus infection and in patients with chronic fatigue syndrome. *J Infect Dis* 1992; **165**: 994–1000.
- 80 Callan MFC, Tan L, Annels N, et al. Direct visualization of antigen-specific CD8⁺ T cells during the primary immune response to Epstein-Barr virus in vivo. *J Exp Med* 1998; **187**: 1395–402.
- 81 Kimura H, Hoshino Y, Kanegane H, et al. Clinical and virologic characteristics of chronic active Epstein-Barr virus infection. *Blood* 2001; **98**: 280–86.
- 82 Straus SE. The chronic mononucleosis syndrome. *J Infect Dis* 1988; **157**: 405–12.
- 83 Okamura T, Hatsukawa Y, Arai H, Inoue M, Kawa K. Blood stem-cell transplantation for chronic active Epstein-Barr virus with lymphoproliferation. *Lancet* 2000; **356**: 223–24.
- 84 Kuzushima K, Yamamoto M, Kimura H, et al. Establishment of anti-Epstein-Barr virus (EBV) cellular immunity by adoptive transfer of virus-specific cytotoxic T lymphocytes from an HLA-matched sibling to a patient with severe chronic active EBV infection. *Clin Exp Immunol* 1996; **103**: 192–98.
- 85 Bar RS, DeLor CJ, Clausen KP, et al. Fatal infectious mononucleosis in a family. *N Engl J Med* 1974; **290**: 363–67.
- 86 Purtillo DT, Cassel CK, Yang JPS, et al. X-linked recessive progressive combined variable immunodeficiency (Duncan's Disease). *Lancet* 1975; **1**: 935–41.
- 87 Mroczek EC, Weisenburger DD, Grierson HL, Markin R, Purtillo DT. Fatal infectious mononucleosis and virus-associated hemophagocytic syndrome. *Arch Pathol Lab Med* 1987; **111**: 530–35.
- 88 Seemayer TA, Gross TG, Egeler RM, et al. X-linked lymphoproliferative disease: twenty-five years after the discovery. *Pediatr Res* 1995; **38**: 471–78.
- 89 Hamilton JK, Paquin LA, Sullivan JL, et al. Finkelstein GZ, landing B, Grunnet M, Purtillo DT. X-linked lymphoproliferative syndrome registry report. *J Pediatr* 1980; **96**: 669–73.
- 90 Arkwright PD, Makin G, Will AM, et al. X linked lymphoproliferative disease in a United Kingdom family. *Arch Dis Child* 1988; **79**: 52–55.
- 91 Migliorati R, Castaldo A, Russo S, et al. Treatment of EBV-induced lymphoproliferative disorder with epipodophyllotoxin VP16-213. *Acta Paediatr* 1994; **83**: 1322–25.
- 92 Gross TG, Filipovich AH, Conley ME, et al. Cure of X-linked lymphoproliferative disease (XLP) with allogeneic hematopoietic stem cell transplantation (HSCT): report from the XLP registry. *Bone Marrow Transplant* 1996; **17**: 741–44.
- 93 Coffey AJ, Brooksbank RA, Brandau O, et al. Host response to EBV infection in X-linked lymphoproliferative disease results from mutations in an SH2-domain encoding gene. *Nat Genet* 1998; **20**: 129–35.
- 94 Sayos J, Wu C, Morra M, et al. Notarangelo L, Geha R, Roncarolo MG, Oetting H, De Vries JE, Aversa G, Terhorst C. The X-linked lymphoproliferative-disease gene product SAP regulates signals induced through the co-receptor SLAM. *Nature* 1998; **395**: 462–69.
- 95 Nichols KE, Harkin DP, Levitz S, et al. Inactivating mutations in an SH2 domain-encoding gene in X-linked lymphoproliferative syndrome. *Proc Natl Acad Sci USA* 1998; **95**: 13765–70.
- 96 Sumegi J, Huang D, Lanyi A, et al. Correlation of mutations of the SH2D1A gene and Epstein-Barr virus infection with clinical phenotype and outcome in X-linked lymphoproliferative disease. *Blood* 2000; **96**: 3118–25.
- 97 Morra M, Silander O, Calpe S, et al. Alterations of the X-linked lymphoproliferative disease gene SH2D1A in common variable immunodeficiency syndrome. *Blood* 2001; **98**: 1321–25.
- 98 Nistala K, Gilmour KC, Cranston T, Davies EG, Goldblatt D, Gaspar, Jones AM. X-linked lymphoproliferative disease: three atypical cases. *Clin Exp Immunol* 2001; **126**: 126–30.
- 99 Dutz JP, Benoit L, Wang X, et al. Lymphocytic vasculitis in X-linked lymphoproliferative disease. *Blood* 2001; **97**: 95–100.
- 100 Brandau O, Schuster V, Weiss M, et al. Epstein-Barr virus-negative boys with non-Hodgkin lymphoma are mutated in the SH2D1A gene, as are patients with X-linked lymphoproliferative disease (XLP). *Hum Mol Genet* 1999; **8**: 2407–13.
- 101 Cocks BG, Chang C-CJ, Carballido JM, Yssel H, de Vries, Aversa G. A novel receptor involved in T-cell activation. *Nature* 1995; **376**: 260–63.
- 102 Tangye SG, Phillips JH, Lanier LL, Nichols KE. Functional requirement for SAP in 2B4-mediated activation of human natural killer cells as revealed by the X-linked lymphoproliferative syndrome. *J Immunol* 2000; **165**: 2932–36.
- 103 Gaspar HB, Sharifi R, Gilmour KC, Thrasher AJ. X-linked lymphoproliferative disease: clinical, diagnostic and molecular perspective. *Br J Haematol* 2002; **119**: 585–95.
- 104 Savoie A, Perpète C, Carpentier L, Joncas J, Alfieri C. Direct correlation between the load of Epstein-Barr virus-infected lymphocytes in the peripheral blood of pediatric transplant patients and risk of lymphoproliferative disease. *Blood* 1994; **83**: 2715–22.
- 105 Lucas KG, Small TN, Heller G, Dupont B. The development of cellular immunity to Epstein-Barr virus after allogeneic bone marrow transplantation. *Blood* 1996; **87**: 2594–603.
- 106 Nalesnik MA. Clinical and pathological features of post-transplant lymphoproliferative disorders (PTLD). *Springer Semin Immunopathol* 1998; **20**: 325–42.
- 107 Ho M, Jaffe R, Miller G, et al. The frequency of Epstein-Barr virus infection and associated lymphoproliferative disease after transplantation and its manifestations in children. *Transplantation* 1988; **45**: 719–27.
- 108 Ho M, Miller G, Atchison RW, et al. Epstein-Barr virus infections and DNA hybridization studies in posttransplantation lymphoma and lymphoproliferative lesions: the role of primary infection. *J Infect Dis* 1985; **152**: 876–86.
- 109 Stevens SJC, Middeldorp JM. Nucleic acid-based diagnostic approaches in EBV-associated disease. *EBV Rep* 2001; **8**: 31–40.
- 110 Lucas KG, Filo R, Heilman DK, Lee CH, Emanuel DJ. Semi-quantitative Epstein-Barr virus polymerase chain reaction analysis of peripheral blood from organ transplant patients and risk for the development of lymphoproliferative disease. *Blood* 1998; **92**: 3977–78.
- 111 Kimura H, Morita M, Yabuta Y, et al. Quantitative analysis of Epstein-Barr virus load by using a real-time PCR assay. *J Clin Microbiol* 1999; **37**: 132–36.
- 112 Baldanti F, Grossi P, Furione M, et al. High levels of Epstein-Barr virus DNA in blood of solid-organ transplant recipients and their value in predicting post-transplant lymphoproliferative disorders. *J Clin Microbiol* 2000; **38**: 613–19.
- 113 Hopwood PA, Brooks L, Parratt R, et al. Persistent Epstein-Barr virus infection: unrestricted latent and lytic viral gene expression in healthy immunosuppressed transplant recipients. *Transplantation* 2002; **74**: 194–202.
- 114 Nalesnik MA, Locker J, Jaffe R, et al. Clonal characteristics of posttransplant lymphoproliferative disorders. *Transplant Proc* 1988; **20**: 280–83.
- 115 Timms JM, Bell A, Flavell F, et al. Epstein-Barr virus (EBV)-positive post transplant lymphoproliferative disease frequently arises from B cells with randomly mutated or sterile immunoglobulin genes: a mechanistic link with EBV-positive Hodgkin's Disease. *Lancet* 2003; **361**: 217–23.
- 116 Starzl TE, Porter KA, Iwatsuki S, Rosenthal JT, Shaw Jr, Atchison RW, Nalesnik MA, Ho M, Griffith BP, Hakala TR, Hardesty RL, Jaffe R, Bahnson HT. Reversibility of lymphomas and lymphoproliferative lesions developing under cyclosporin-steroid therapy. *Lancet* 1984; **1**: 583–87.
- 117 Armitage JM, Kormos RL, Stuart RS, et al. Posttransplant lymphoproliferative disease in thoracic organ transplant patients: ten years of cyclosporine-based immunosuppression. *J Heart Lung Transplant* 1991; **10**: 877–87.
- 118 Roche, MabThera. http://www.rocheuk.com/ProductDB/Documents/rx/spc/Mabthera_100mg_10ml_SPC.pdf.
- 119 Cook RC, Connors JM, Gascoyne RD, Fradet G, Levy RD. Treatment of post-transplant lymphoproliferative disease with rituximab monoclonal antibody after lung transplantation. *Lancet* 1999; **354**: 1698–99.
- 120 Kuehnle I, Huls MH, Liu Z, et al. CD20 monoclonal antibody (rituximab) for therapy of Epstein-Barr virus lymphoma after hemopoietic stem-cell transplantation. *Blood* 2000; **95**: 1502–05.
- 121 Verschuuren EAM, Stevens SJC, van Imhoff GW, Middeldorp JM, Boer C de, Koëter G, The TH, van der Bij W. Treatment of posttransplant lymphoproliferative disease with rituximab: the remission, the relapse and the complication. *Transplantation* 2002; **73**: 100–04.
- 122 Anon. Rituximab (MabThera): serious infusion-related adverse reactions. *Curr Probl Pharmacovigilance* 1999; **25**: 2–3.
- 123 Suzan F, Ammor M, Ribrag V. Fatal reactivation of cytomegalovirus infection after use of rituximab for a post-transplantation lymphoproliferative disorder. *N Engl J Med* 2001; **13**: 1000.
- 124 Sharma VR, Fleming DR, Slone SP. Pure red cell aplasia due to parvovirus B19 in a patient treated with rituximab. *Blood* 2000; **96**: 1184–86.
- 125 Rooney CM, Smith CA, Ng CYC, et al. Infusion of cytotoxic T cells for the prevention and treatment of Epstein-Barr virus-induced lymphoma in allogeneic transplant recipients. *Blood* 1998; **92**: 1549–55.
- 126 Haque T, Wilkie GM, Taylor C, et al. Treatment of Epstein-Barr virus-positive post-transplantation

- lymphoproliferative disease with partly HLA-matched allogeneic cytotoxic T cells. *Lancet* 2002; **360**: 436–42.
- 127 Herbst H. Epstein-Barr virus in Hodgkin's disease. *Semin Cancer Biol* 1996; **7**: 183–89.
 - 128 Mueller N, Evans A, Harris NL, et al. Hodgkin's disease and Epstein-Barr virus. Altered antibody pattern before diagnosis. *N Engl J Med* 1989; **320**: 689–95.
 - 129 Gutensohn N, Cole P. Epidemiology of Hodgkin's disease. *Semin Oncol* 1980; **7**: 92–102.
 - 130 Weiss LM, Movahed LA, Warnke RA, Sklar J. Detection of Epstein-Barr viral genomes in Reed-Sternberg cells of Hodgkin's disease. *N Engl J Med* 1989; **320**: 502–06.
 - 131 Jarrett RF. Viruses and Hodgkin's lymphoma. *Ann Oncol* 2002; **13**: 23–29.
 - 132 Pallesen G, Hamilton-Dutoit SJ, Rowe M, Young LS. Expression of Epstein-Barr virus latent gene products in tumour cells of Hodgkin's disease. *Lancet* 1991; **337**: 320–22.
 - 133 Young LS, Dawson CW, Eliopoulos AG. The expression and function of Epstein-Barr virus encoded latent genes. *J Clin Pathol: Molecular Pathology* 2000; **53**: 238–47.
 - 134 Anagnostopoulos I, Herbst H, Niedobitek G, Stein H. Demonstration of monoclonal EBV genomes in Hodgkin's disease and KI-1 positive anaplastic large cell lymphoma by combined Southern blot and in situ hybridization. *Blood* 1989; **74**: 810–16.
 - 135 Marafioti T, Hummel M, Foss H-D, et al. Hodgkin and Reed-Sternberg cells represent an expansion of a single clone originating from a germinal centre B-cell with functional immunoglobulin gene rearrangements but defective immunoglobulin transcription. *Blood* 2000; **95**: 1443–50.
 - 136 Lukes RJ, Tindle BH, Parker JW. Reed-Sternberg-like cells in infectious mononucleosis. *Lancet* 1969; **2**: 1003–04.
 - 137 Chetty R, Biddolph S, Gatter K. An immunohistochemical analysis of Reed-Sternberg-like cells in posttransplantation lymphoproliferative disorders: the possible pathogenetic relationship to Reed-Sternberg cells in Hodgkin's disease and Reed-Sternberg-like cells in Non-Hodgkin's lymphomas and reactive conditions. *Hum Pathol* 1997; **28**: 493–98.
 - 138 Rickinson AB, Moss DJ. Human cytotoxic T lymphocyte responses to Epstein-Barr virus infection. *Ann Rev Immunol* 1997; **15**: 405–31.
 - 139 Roskrow MA, Suzuki N, Gan Y-J, et al. Epstein-Barr virus (EBV)-specific cytotoxic T lymphocytes for the treatment of patients with EBV-positive relapsed Hodgkin's disease. *Blood* 1998; **91**: 2925–34.
 - 140 Baarle D van, Hovenkamp E, Callan MFC, et al. Dysfunctional Epstein-Barr virus (EBV)-specific CD8+ T lymphocytes and increased EBV load in HIV-1 infected individuals progressing to AIDS-related non-Hodgkin lymphoma. *Blood* 2001; **98**: 146–55.
 - 141 Beral V, Peterman T, Berkelman R, Jaffe H. AIDS-associated non-Hodgkin Lymphoma. *Lancet* 1991; **337**: 805–09.
 - 142 MacMahon EME, Glass JD, Hayward SD, Mann RB. Epstein-Barr virus in AIDS-related primary central nervous system lymphoma. *Lancet* 1991; **338**: 969–73.
 - 143 Cinque P, Brytting M, Vago L, et al. Epstein-Barr virus DNA in cerebrospinal fluid from patients with AIDS-related lymphoma of the central nervous system. *Lancet* 1993; **342**: 398–401.
 - 144 Kirk O, Pedersen C, Cozzi-Lepri A, et al. Non-Hodgkin lymphoma in HIV-infected patients in the era of highly active antiretroviral therapy. *Blood* 2001; **98**: 3406–12.
 - 145 Hoffman C, Tabrizian S, Wolf E, et al. Survival of AIDS patients with primary central nervous system lymphoma is dramatically improved by HAART-induced immune recovery. *AIDS* 2001; **15**: 2119–27.
 - 146 Powles T, Bower M. HIV-associated Hodgkin's disease. *Int J STD AIDS* 2000; **11**: 492–94.
 - 147 Grulich AE, Li Y, McDonald A, et al. Rates of non-AIDS-defining cancers in people with HIV infection before and after AIDS diagnosis. *AIDS* 2002; **16**: 1155–61.
 - 148 Greenspan JS, Greenspan D, Lennette ET, et al. Replication of Epstein-Barr virus within the epithelial cells of oral hairy leukoplakia: virus replication in the absence of a detectable latent phase. *N Engl J Med* 1985; **313**: 1564–71.
 - 149 Itin P, Ruffli T, Rudlinger R, et al. Oral hairy leukoplakia in a HIV-negative renal transplant patient: a marker for immunosuppression? *Dermatologica* 1988; **177**: 126–28.
 - 150 Greenspan D, Greenspan JS, Souze YG De, Levy JA, Ungar AM. Oral hairy leukoplakia in an HIV-negative renal transplant recipient. *Oral Pathol* 1989; **18**: 32–34.
 - 151 Flückiger R, Laifer G, Itin P, Meyer B, Lang C. Oral hairy leukoplakia in a patient with ulcerative colitis. *Gastroenterology* 1994; **106**: 506–08.
 - 152 Niedobitek G, Young LS, Lau R, et al. Epstein-Barr virus infection in oral hairy leukoplakia: virus replication in the absence of a detectable latent phase. *J Gen Virol* 1991; **72**: 3035–46.
 - 153 Thomas JA, Felix DH, Wray D, et al. Epstein-Barr virus gene expression and epithelial cell differentiation in oral hairy leukoplakia. *Am J Pathol* 1991; **139**: 1369–80.
 - 154 Resnick L, Herbst JS, Ablashi DV, et al. Regression of oral hairy leukoplakia after orally administered acyclovir therapy. *JAMA* 1988; **259**: 384–88.
 - 155 Thorley-Lawson DA, Poodry CA. Identification and isolation of the main component (gp350-gp220) of Epstein-Barr virus responsible for generating neutralising antibodies in vivo. *J Virol* 1982; **43**: 730–36.
 - 156 Khanna R, Sherritt M, Burrows SR. EBV structural antigens, gp350 and gp85, as targets for ex vivo virus-specific CTL during acute infectious mononucleosis: potential use of gp350/gp85 CTL epitopes for vaccine design. *J Immunol* 1999; **162**: 3063–69.
 - 157 Airey D, Le T, Malliaros J, et al. Analysis of EBV polytope ISCOM[®] vaccines for the induction of CTL (abstr). 10th Biennial Conference on EBV and Associated Diseases; Cairns: Australia, 16–21 July 2002.
 - 158 Silins SL, Sherritt MA, Silleri JM, et al. Asymptomatic primary Epstein-Barr virus infection occurs in the absence of blood T-cell repertoire perturbations despite high levels of systemic viral load. *Blood* 2001; **98**: 3739–44.
 - 159 Wang F. A new animal model for Epstein-Barr virus pathogenesis. *Curr Top Microbiol Immunol* 2001; **258**: 201–19.